



EFFECT OF CARBONATES ON SEED GERMINATION AND SEEDLING GROWTH IN *PORTULACA OLERACEA* L.

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ABSTRACT

One of the most important stresses is alkali stress that effects plants growth and metabolism. The effect of carbonates (NaHCO_3 , KHCO_3 and $(\text{NH}_4)_2\text{CO}_3$) on seed germination and seedling growth in *Portulaca oleracea* L. was investigated. Germination percentage was decrease with increasing concentration of salts. Root length and dry weight were strongly reduced due to carbonate salts. Shoot length and fresh weight were slightly increased at lower concentration (25mM) of salts and then decreased at higher concentrations (50, 100, and 200mM). From the results, it appears that *P. oleracea* L. is rather sensitive to carbonate salts during germination and seedling growth.

Keywords: Seed germination, carbonates, alkalinity stress and *Portulaca oleracea* L.

1. INTRODUCTION

Salinisation of soil has become global environmental problem an important factor limiting agricultural productivity. In saline environment adaptation of plants to salinity during germination and early seedling stage is crucial for establishment of species [1]. Salt stress decreases growth in most plants including halophytes. When a saline soil contains HCO_3^- and/or CO_3^{2-} which elevate soil pH, plant experiences damages from both salt and alkali stress [2].

The high pH environment that surrounds the roots can cause metal ions and phosphorus to precipitate [3]. The problem of alkalization due to NaHCO_3 and Na_2CO_3 may be more severe than the problem of salinization caused by the neutral salts such as NaCl and Na_2SO_4 [4].

Salinity impairs seed germination, reduces nodule formation, retard plant development and reduces crop yield [5]. Germination and seedling characteristics are the most viable criteria used for determining salt tolerance in plants [6]. Seed germination and seedling growth are reduced in saline soil with varying responses for species and cultivar [7]. It is usually the most critical stage in seedling establishment [5, 6]. Salinity stress negatively affects seed germination, either osmotically through the accumulation of sodium and chloride causing an imbalance in nutrient uptake or the toxicity effects [7]. Seed germination, seedling emergence and its survival are particularly sensitive to substrate salinity [8, 9].

Portulaca oleracea (Purselane) is an annual plant from portulacaceae [10]. It is the main weed for 45 cultivating types in 81 countries of world [11]. Life cycle of this plant is

completed during 2 to 4 months [12]. The aim of the present study was to evaluate the effects of carbonates (NaHCO_3 , KHCO_3 and $(\text{NH}_4)_2\text{CO}_3$) on germination and early seedling growth.

2. MATERIAL AND METHODS

Seeds of *Portulaca oleracea* L. were collected from the fields of village Kaulage of Kolhapur district. They were surface sterilized with 0.5% sodium hypochloride for 1 min, washed first with a jet of tap water and then with sterile distilled water and air dried. The air dried seeds were kept in a sterilized petridish for germination using germination paper, moistened with 5ml of distilled water or with treatment solutions [25, 50, 100, 200mM concentrations of each NaHCO_3 , KHCO_3 , and $(\text{NH}_4)_2\text{CO}_3$]. The petridishes were kept in the dark for germination. Each treatment had three replicates. Germination percentage was measured after every 24 hours for five days and root length, shoot length and fresh and dry weights of seedlings were measured after 120 h.

3. RESULTS AND DISCUSSIONS

Effects of carbonates on germination and seedling growth (Germination percentage, root length, shoot length, fresh and dry weights) have been depicted in Figures 1 to 5.

It was found that the germination percentage was significantly reduced in all treatments as compared to that in control. Seed germination was totally inhibited at higher concentrations of all carbonates. It was decreased with the increase in NaHCO_3 , KHCO_3 and $(\text{NH}_4)_2\text{CO}_3$ salts concentrations. Only $16.66\% \pm 2.887$ seeds were germinated

at 25 mM $(\text{NH}_4)_2\text{CO}_3$, whereas $32.33\% \pm 3.055$ seeds in KHCO_3 and $21.67\% \pm 11.547$ seeds in NaHCO_3 were germinated. Germination percentage was decreased in all treatments in the following order $\text{Control} \geq \text{KHCO}_3 \geq \text{NaHCO}_3 \geq (\text{NH}_4)_2\text{CO}_3$.

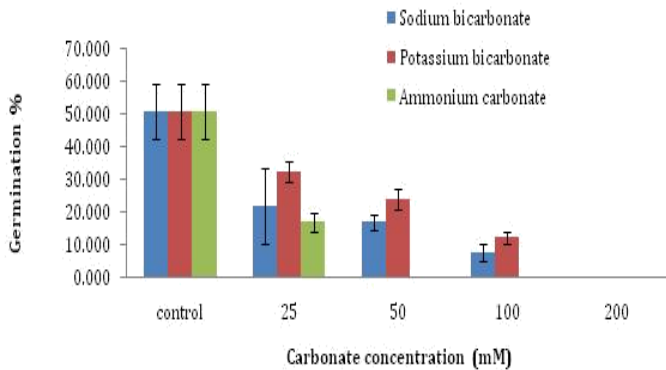


Figure 1: Effect of carbonates on germination percentage in *Portulaca oleracea* L

Root length was decreased with increasing in concentration of carbonate salts. Highest root length was recorded in control ($1.120 \pm 0.253\text{cm}$). Maximum decrease in the root length was observed in KHCO_3 (0.199 ± 0.088) at 50 mM, 0.420 ± 0.123 cm in $(\text{NH}_4)_2\text{CO}_3$ and 0.389 ± 0.078 cm in NaHCO_3 at 50 mM concentration.

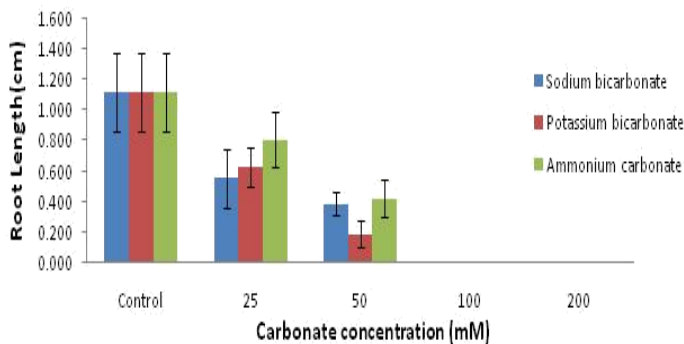


Figure 2: Effect of carbonates on root length of *Portulaca oleracea* L. seedlings.

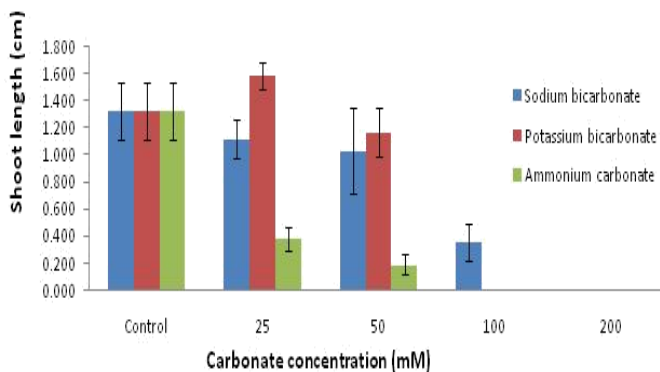


Figure 3: Effect of carbonates on shoot length of *Portulaca oleracea* L. seedlings

Higher shoot length of seedlings was observed in KHCO_3 at the lower concentration (25 mM $1.595 \pm 0.099\text{cm}$) than that in control ($1.322 \pm 0.211\text{cm}$). It was significantly decreased due to NaHCO_3 and $(\text{NH}_4)_2\text{CO}_3$ salinities. It was severely affected by $(\text{NH}_4)_2\text{CO}_3$ ($0.190 \pm 0.074\text{cm}$) as compared to that by NaHCO_3 ($1.033 \pm 0.320\text{cm}$) and KHCO_3 ($1.170 \pm 0.177\text{cm}$) at 50 mM salt concentration. The negative effect of carbonate salts on shoot length of this plant can be given in the order $\text{KHCO}_3 \leq \text{NaHCO}_3 \leq (\text{NH}_4)_2\text{CO}_3$.

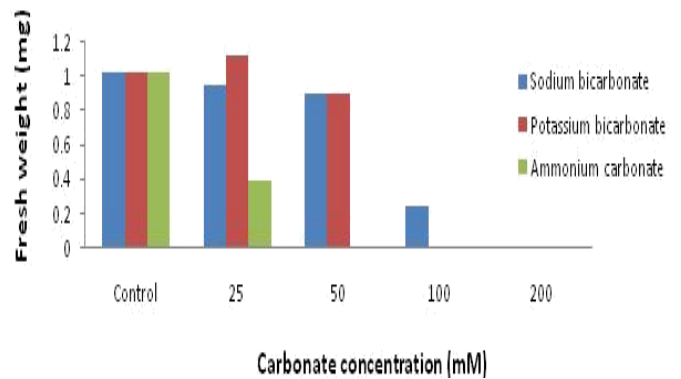


Figure 4: Effect of carbonates on fresh weight of *Portulaca oleracea* L. seedlings

Fresh weight of *Portulaca* seedlings was decreased with increase in the concentration of carbonate except KHCO_3 . At 25 mM KHCO_3 the seedlings fresh weight was slightly increased (1.13 mg) over the control (1.03mg). Dry weight of seedlings was not significantly decreases in lower concentration of $(\text{NH}_4)_2\text{CO}_3$ and KHCO_3 , but at higher concentration of salts (50 and 100 mM) it was negligible.

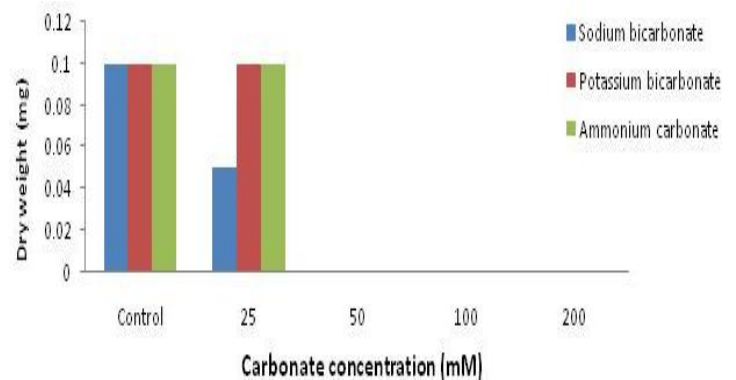


Figure 5: Effect of carbonates on dry weight of *Portulaca oleracea* L. seedlings.

4. DISCUSSION

Salinity is a major environmental stress factor that affects seed germination in coastal marshes [13]. Seed germination of *C. glaucum* was decreased with increase in NaCl , Na_2CO_3 , Na_2SO_4 , MgSO_4 and MgCl concentration in medium [14]. Germination percentage in Thyme (*Thymus vulgaris* L.) seeds

was found decreased at all salinity levels [15]. The salinity stress affects germination in a variety plants which can infer to meaningful decreasing on *Vigna radiate* L. and *Lens culanuris* [16]. Salt stress inhibited the seedling growth and root length was more affected than shoot length [17]. Germination percentage of *Portulaca oleracea* L. was decreased in all imposed salinity levels and the highest germination percentage was observed in control [18]. The dry mass of leaves, stems and roots of sunflower seedlings was decreased with increasing stress intensity; the reduction under alkaline conditions was greater than that under saline conditions [19]. Seedling emergence, fresh and dry mass of both shoot and root as well as seed yield were decreased with increasing salinity [20]. No seed was germinated at the concentration of 250mM NaCl in case of *Portulaca oleracea* L [21]. The fresh weight of germinated seed was significantly modified, under the influence of salt stress at different levels in *Vigna unguiculata* Cv. Pitiuba [22].

5. REFERENCES

- Lianes A, Reinoso H, Luna V, *World J. of Agricultural Sciences*, 2005; **1**(2), 120-128.
- Munns R, *Plant Cell Environ*, 2002; **25**:239-250.
- Shi DC, Zhao KF, *Acta Pratacu. Sin*, 1997; **6**:51-61.
- Kawanabe S, and Zhu TC, *J. Jpn. Grassl. Sci.*, 1991; **37**:91-99.
- Greenway H, Munns R, *Annu. Rev. Plant Physiol*, 1980; **31**:149-190.
- Almansouri MJ, Kinet M, Lutts S., *Plant and Soil*, 2001; **231**: 243-254.
- Bliss RD, Platt-Aloia KA, Thompson WW, *Plant, Cell and Environment*, 1986; **9**:721-726.
- Maiko S, Kachi N, Ishikawa S, Furukawa A, *Ecology Res*, 1992; **7**: 225-233.
- Baldwin H, Mckee KL, Mendelssohn IA, *Amer J Bot*, 1996; **83**: 470-479.
- Schuman M, *NNFA Today*, 2001; **15**(6):12.
- Holm LG, Plucknett DL, Pancho JV, Herberger JP, *University of Hawaii Press, Honolulu, Hawaii USA*, 1997.
- Chauhan BS, Jonson DE, *Crop and Environmental Sciences Division, IRRI*, 2007.
- Khan MA, International symposium on optimum resource utilisation in salt affected ecosystem in arid and semiarid regions (Eds.): Desert research center, Cario, Egypt, Pp.346-358.
- Duan D, Liu X, Khan MA, Gul B, *Pak. J. Bot*, 2004; **36**(4):793-800.
- Hoseini MS, *IJACS*, 2010; **2-1**:34-38.
- Kazerouni E, Akramian M, Tokassi S, and Eghbali S. International symposium on optimum resource utilization in salt affected ecosystems in arid and semi arid regions (Eds.):Desert Research Centre, Cairo, Egypt. 2005; pp. 346-358.
- Jamil MD, Lee KY, Jung M, Ashraf SC, et al., *J. Cent. Eur. Agric*, 2006; **7**:273-282.
- Rahbari P, Tavakoli S, Hosseini SM, *J. of stress physiology and biochemistry*, 2011; **8**:183-193.
- Liu J, Guo wQ, Shi DC, *Photosynthetica*, 2010; **48**(2):278-286.
- Mensah JK, Ihenyen J, *Nigerian annals Of natural science*. 2009; **8**(2):17-24.
- Chauhan BS, Johnson DE, *Ann Appl Biol*, 2009; **155**:61-69.
- Labato AKS, Santos Filho BG, Costa RCL, Goncalves- Vidigal MC et al., *World J. of Agri Sci*, 2009; **5**(5):590-596.