



## DEVELOPMENT AND STANDARDIZATION OF POLYHERBAL FORMULATION

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## ABSTRACT

The most important challenges posed by herbal formulations is their evaluation and standardization. Evaluation is necessary to ensure the quality and purity of the herbal product. For evaluation of raw materials and poly herbal formulation various parameters are studied as per the World Health Organization's guidelines and the Ayurvedic Pharmacopoeia of India. The present study deals with formulation the polyherbal formulation prepared from hydro-alcoholic (30:70) extracts of *Andrographis paniculata* (Stem and leaves), *Asparagus racemosus* (Root), *Ipomoea digitata* (Rhizome), *Tinospora cordifolia* (Stem) and *Withania somnifera* (Root). To evaluate of raw materials include physicochemical studies like ash values, extractive values, phytochemical studies and safety profiles which include heavy metal analysis, pesticide residue analysis and microbial load analysis. The Preformulation parameters and parameters for finished product (hard gelatin capsule) include uniformity of weight, disintegration time, moisture content, pH, phytochemical estimation and microbial load assay. The HPTLC finger print profile of finished product was also carried out.

**Keywords:** Development, Standardization, Polyherbal formulation, WHO guidelines

## 1. INTRODUCTION

Plants are very useful to mankind. Many of them are used exclusively for medicinal purposes. According to the World Health Organization (WHO), "a medicinal plant is a plant which, in one or more of its organs, contains substances that can be used for therapeutic purposes, or which are precursors for chemo-pharmaceutical semi-synthesis." Such plants are in great demand by pharmaceutical companies for their active ingredients [1, 2].

Diabetes mellitus (DM) is a chronic complication of derangement of protein, carbohydrate and lipid metabolism characterized by increased blood glucose level resulting from the defects in insulin secretion, insulin action or both [3]. DM is the worldwide problem to leading micro vascular and macrovascular complications [4]. DM is a chronic complication that affected an estimated 135 million people in 1995, 285 million people worldwide in 2010 and data reached that approx 500 million people in 2025 mainly in rural and poor population throughout the world [5].

In traditional systems of medicine, many plants have been documented to be useful for the treatment of various systemic disorders. Many of the traditional/indigenous systems of medicine are effective but they suffer from lack of complete standardization which is one of the important challenges posed by the traditional systems of medicine. The concept of polyherbal formulation is well documented in the ancient literature. Compared to the single herb, the polyherbal formulation has better and extended therapeutic potential. Hence, the present study was planned to formulate and

standardize a polyherbal formulation using plants having known anti diabetic potential.

## 2. MATERIALS AND METHODS

## 2.1. Collection of the plant and Authentication

The selected plant materials viz *Andrographis paniculata* (Stem and leaves), *Asparagus racemosus* (Root), *Ipomoea digitata* (Rhizome), *Tinospora cordifolia* (Stem), *Withania somnifera* (Root) were procured from the authentic suppliers and further authenticated by Dr. P. Jayaraman, Director, Plant Anatomy and Research Centre, Tambaram, Chennai. (Specimen numbers PARC/2015/3049 for *Andrographis paniculata*, PARC/2015/3053 for *Asparagus racemosus*, PARC/2015/3050 for *Ipomoea digitata*, PARC/2015/3052 for *Tinospora cordifolia*, PARC/2015/3051 for *Withania somnifera*).

## 2.2. Standardization of raw material

The Organoleptic evaluation and determination of Foreign organic matter of raw materials were carried out as per the Ayurvedic Pharmacopoeia of India [6].

## 2.3. Physicochemical studies

The ash values, extractive values and loss on drying were performed according to the official methods prescribed in Ayurvedic Pharmacopoeia of India [6].

## 2.4. Preliminary phyto-chemical screening

One gram of each of the extracts of *Andrographis paniculata* (Stem and leaves), *Asparagus racemosus* (Root), *Ipomoea digitata* (Rhizome), *Tinospora cordifolia* (Stem),

*Withania somnifera* was dissolved in 100 ml of its own mother solvent to obtain a stock of concentration 1% w/v and tested for the presence of alkaloids, flavanoid, tannins, saponin, glycosides, terpenoids, steroid, carbohydrate and phenolic compounds, and saponins [7].

## 2.5. Fluorescence analysis

The crude drug was studied for any fluorescence under ordinary light and UV light. Samples were studied as such, before and after treating with 50% HCl and 50% NaOH and the results were tabulated [8].

## 2.6. Safety profile studies

The safety profile parameters like heavy metal analysis, pesticide residual analysis and microbial load analysis were studied according to the official methods prescribed in Ayurvedic Pharmacopoeia of India.

### 2.6.1. Quantitative estimation of heavy metals

Quantitative estimation of heavy metals was done for the detection of arsenic, lead, cadmium and mercury as per the Ayurvedic pharmacopoeial procedures [9].

### 2.6.2. Quantitative estimation of pesticide residues

Quantitative estimation of pesticide were done for the detection of Organochlorine compound, Organophosphorous compound and Carbamates as per the Ayurvedic pharmacopoeial procedures [10].

### 2.6.3. Microbial load analysis

For the safe use of raw material for the polyherbal capsules, microbial count was done and it was checked whether the total aerobic viable count, yeasts and molds were within the prescribed limits and the microorganisms, *Escherichia coli*, *Salmonellae*, *Pseudomonas Staphylococcus* and *Shigella* were checked [11].

## 2.7. Preparation of Extract

The selected plants materials were shade dried and stored in an airtight container, each drug coarsely powdered and extracted with hydro-alcoholic (30:70) by hot percolation method using soxhlet apparatus. The prepared hydro-alcohol extracts were concentrated under vacuum using rotary vacuum evaporator at 40°C temperature (removal of alcohol). The concentrated extracts were freeze dried at -20°C. The powders were stored in an airtight container and kept in the desiccator until further use.

## 2.8. Formula of Polyherbal formulation

The polyherbal formulation (capsules) contained the hydro-alcoholic extracts of *Andrographis paniculata*, *Asparagus racemosus*, *Ipomoea digitata*, *Tinospora cordifolia*, *Withania somnifera* in the ratio of 1:1:1:1:1.

## 2.9. Preparation of formulation by wet granulation method

The formulation preparation began with trials by adding a different ratio of binders and selecting the quantity of

lubricants and preservatives, and finally the procedure was optimized. *Andrographis paniculata*, *Asparagus racemosus*, *Ipomoea digitata*, *Tinospora cordifolia*, *Withania somnifera* extracts were powdered (sieve 40), and mixed in the ratio of 1:1:1:1:1 and taken for the preparation of capsules by wet granulation technique using 5% starch paste as a binder. The wet mass was passed through sieve number 22 to obtain granules. The granules were dried at 45°C in a tray [12].

## 2.10. Pre formulation studies.

Preformulation parameters such as bulk density, tap density, Compressibility index, Hausner's ratio, and angle of repose were determined for the prepared polyherbal granules and the best trial batch were taken for capsule filling and further studies [13, 14].

## 2.11. Standardization of polyherbal formulation (hard gelatin capsule)

**Capsule evaluation:** The polyherbal capsules were evaluated for their description, average weight, weight variation, moisture content, disintegration time, pH and microbial load and compared with Indian pharmacopoeial standards<sup>[15]</sup>. and also preliminary screening of phyto constituents, quantitative estimation of phyto-constituent were carried out.

**Average weight:** Twenty capsules were individually weighed and the average weight of the capsule was calculated.

**Weight variation:** The individual weights of the each capsule should be within the limits of 90% and 110% of the average weight.

**Moisture content:** Moisture content was determined by using automatic Karl Fischer titration apparatus.

**Disintegration time:** Disintegration test was performed using the digital microprocessor based disintegration test apparatus (Electro lab, Mumbai, India). One capsule was introduced into each tube and a disc was added to each tube. The assembly was suspended in water in a 1000 ml beaker. The volume of water at its highest point was at least 25 mm below the surface of the water and at its lowest point was at least 25 mm above the bottom of the beaker. The apparatus was operated and maintained at a temperature of  $37 \pm 2^\circ\text{C}$ .

**pH value:** pH of 1% solution was determined by using a digital pH meter.

### 2.11.1. Preliminary phyto-chemical screening

One gram of polyherbal formulation was dissolved in 100 ml of its own mother solvent to obtain a stock of concentration 1% w/v and tested for the presence of alkaloids, flavanoid, tannins, saponin, glycosides, terpenoids, steroid, carbohydrate and phenolic compounds, and saponins [7].

### 2.11.2. Fluorescence analysis

The crude drug was studied for any color changes under ordinary light and UV light. Samples were studied as such, after treating with 50% HCl and 50% NaOH and the results were tabulated [8].

### 2.11.3. Quantitative estimation of phytoconstituents

Total Alkaloids, Phenolics compounds, Flavonoid and Tannin Content were estimated in the polyherbal formulation [16-19].

### 2.11.4. Microbial load analysis

For the safe use of raw material for the polyherbal capsules, microbial count was done and it was checked whether the total aerobic viable count, yeasts and molds were within the prescribed limits and the microorganisms, *Escherichia coli*, *Salmonellae*, *Pseudomonas* *Staphylococcus* and *Shigella* were checked.

### 2.11.5. High-performance thin layer chromatography (HPTLC) fingerprint analysis

Randomly few capsules were selected. They were opened and the contents were collected. The capsule content was dissolved in a minimum volume of mobile phase and used for the fingerprint analysis. Twenty milligram either individual herbal extract or polyherbal formulation was reconstituted in 1 ml of ethanol and filtered through a 0.45  $\mu$ m membrane filter. The filtrate was used for the HPTLC analysis. The solvent system Toluene:Ethylacetate:Methanol:Formicacid (6:3:1:0.2) was used as mobile phase for the development of chromatogram [20, 21].

## 3. RESULTS AND DISCUSSION

The most important part of any formulation is standardization which ensures the quality, safety and reproducibility. It involves the complete process of bio-prospection right from the collection of raw materials to development of finished product. In the present study, standardized polyherbal mixture was formulated in hard gelatin capsule to replace the traditional liquid dosage form.

Polyherbal formulation composed of five ingredients, belonging to different families, different morphological plant parts and different phytoconstituents.

**Table 3: Physicochemical parameters**

| Physicochemical parameters | <i>Andrographis paniculata</i> | <i>Asparagus racemosus</i> | <i>Ipomoea digitata</i> | <i>Tinospora cordifolia</i> | <i>Withania somnifera</i> |
|----------------------------|--------------------------------|----------------------------|-------------------------|-----------------------------|---------------------------|
| Loss on drying             | 2.07 $\pm$ 0.15                | 5.78 $\pm$ 0.25            | 5.84 $\pm$ 0.36         | 4.84 $\pm$ 0.36             | 6.23 $\pm$ 0.25           |
| Total ash value            | 6.58 $\pm$ 0.28                | 3.87 $\pm$ 0.47            | 3.72 $\pm$ 0.28         | 9.52 $\pm$ 0.76             | 6.09 $\pm$ 0.15           |
| Acid insoluble ash         | 1.23 $\pm$ 0.51                | 0.38 $\pm$ 0.08            | 0.72 $\pm$ 0.07         | 1.74 $\pm$ 0.51             | 0.34 $\pm$ 0.02           |
| water soluble ash          | 1.57 $\pm$ 0.23                | 2.37 $\pm$ 0.22            | 1.17 $\pm$ 0.52         | 2.34 $\pm$ 0.42             | 1.23 $\pm$ 0.32           |
| Sulphated ash              | 1.25 $\pm$ 0.55                | 7.59 $\pm$ 0.87            | 9.05 $\pm$ 0.72         | 13.16 $\pm$ 0.57            | 10.08 $\pm$ 0.25          |
| Water soluble Extractive   | 21.74 $\pm$ 0.26               | 53.63 $\pm$ .35            | 19.52 $\pm$ 0.71        | 45.72 $\pm$ 0.86            | 28.76 $\pm$ 0.35          |
| Alcohol soluble extractive | 24.5 $\pm$ 0.86                | 31.28 $\pm$ 0.58           | 25.23 $\pm$ 0.57        | 10.75 $\pm$ 25              | 17.23 $\pm$ 25            |
| Ether soluble extractive   | 10.28 $\pm$ 0.85               | 8.35 $\pm$ 0.53            | 23.68 $\pm$ 0.48        | 8.50 $\pm$ 0.25             | 13.25 $\pm$ 0.84          |

The value are expressed as mean  $\pm$  SD, (n=3); NMT-Not more than

### 3.1. Foreign Organic Matter

Foreign organic matter for the raw materials was done with samples taken from the suppliers. The results obtained and the standard values are given in table 1.

**Table1: Foreign organic matter of samples**

| Plant Name                     | Observation     | Acceptable Limits (w/w %) |
|--------------------------------|-----------------|---------------------------|
| <i>Andrographis paniculata</i> | 0.62 $\pm$ 0.32 | NMT 2                     |
| <i>Asparagus racemosus</i>     | 0.65 $\pm$ 0.37 | NMT 2                     |
| <i>Ipomoea digitata</i>        | 0.85 $\pm$ 0.58 | NMT 2                     |
| <i>Tinospora cordifolia</i>    | 0.76 $\pm$ 0.42 | NMT 2                     |
| <i>Withania somnifera</i>      | 0.51 $\pm$ 0.21 | NMT 2                     |

The value are expressed as mean  $\pm$  SD, (n=3); NMT-Not more than

### 3.2. Organoleptic Characters

Organoleptic characters for the raw materials were done with samples taken from the suppliers. The results obtained and the standard values are given table 2.

**Table 2: Organoleptic characters of selected plants**

| Plant Name                     | Nature        | Colour       | Odour          | Taste          |
|--------------------------------|---------------|--------------|----------------|----------------|
| <i>Andrographis paniculata</i> | Coarse powder | Green        | Characteristic | Bitter         |
| <i>Asparagus racemosus</i>     | Coarse powder | Pale yellow  | Aromatic       | Bitter         |
| <i>Ipomoea digitata</i>        | Coarse powder | White        | Characteristic | Characteristic |
| <i>Tinospora cordifolia</i>    | Coarse powder | Light brown  | Aromatic       | Bitter         |
| <i>Withania somnifera</i>      | Coarse powder | Light yellow | Characteristic | Bitter         |

### 3.3. Physicochemical Parameters

Various physicochemical parameters were calculated for the herbal drugs used in the polyherbal formulation. Table 3 depicts the report of various physicochemical parameters.

### 3.4. Phytochemical Analysis

The chemical tests for various Phyto constituents in the raw materials were carried out and the results were recorded and detailed in Table 4.

**Table 4: Phytochemical Analyses**

| Plant Name                     | Alkaloid | Flavanoid | Tannin | Saponin | Glycoside | Terpenoid | Steroid | Carbohydrate | Phenolics |
|--------------------------------|----------|-----------|--------|---------|-----------|-----------|---------|--------------|-----------|
| <i>Andrographis paniculata</i> | +        | +         | +      | -       | +         | +         | +       | -            | +         |
| <i>Asparagus racemosus</i>     | +        | +         | +      | -       | +         | +         | +       | -            | +         |
| <i>Ipomoea digitata</i>        | +        | +         | -      | -       | +         | -         | -       | +            | -         |
| <i>Tinospora cordifolia</i>    | +        | -         | +      | -       | +         | +         | +       | +            | -         |
| <i>Withania somnifera</i>      | +        | -         | +      | +       | -         | +         | -       | -            | +         |

**3.5. Fluorescence Analysis of Raw Materials****Table 5: Fluorescence Analysis**

| Sample                      | Before Treatment |             |                | After treating with 50 % HCl |                 |            | After treating with 50% NaOH |                 |                    |
|-----------------------------|------------------|-------------|----------------|------------------------------|-----------------|------------|------------------------------|-----------------|--------------------|
|                             | Ordinary light   | Short UV    | Long UV        | Ordinary light               | Short UV        | Long UV    | Ordinary light               | Short UV        | Long UV            |
| <i>A. paniculata</i>        | Greenish brown   | Green       | Green          | Green                        | Greenish brown  | Brown      | Greenish yellow              | Greenish yellow | Dark brown         |
| <i>Asparagus racemosus</i>  | Light brown      | Brown       | Dark brown     | Light brown                  | Brownish green  | Dark brown | Light brown                  | Greenish brown  | Green fluorescence |
| <i>Ipomoea digitata</i>     | Dark brown       | Light brown | Greenish brown | Brown                        | Greenish yellow | Dark brown | Light brown                  | Greenish brown  | Green              |
| <i>Tinospora cordifolia</i> | Dull brown       | Cream       | Brown          | Brown                        | Brownish white  | Dark brown | Light brown                  | Brownish white  | Dark brown         |
| <i>Withania somnifera</i>   | Greenish brown   | Green       | Light brown    | Brown                        | Brown           | Dark brown | Light brown                  | Dull brown      | Light brown        |

**3.6. Safety Profile Parameters Studies****3.6.1. Heavy metal analysis**

Quantitative estimation of heavy metals

Quantitative estimation of heavy metals in the raw materials was carried out and the results were recorded and detailed in Table 6.

**Table: 6 Test for heavy metals**

| Plant name           | OBSERVATION (in ppm/ml) |               |                   |                   |
|----------------------|-------------------------|---------------|-------------------|-------------------|
|                      | Arsenic (NMT 5)         | Lead (NMT 10) | Cadmium (NMT 0.3) | Mercury (NMT 0.5) |
| <i>A. paniculata</i> | 0.008                   | 0.081         | 0.025             | 0.004             |
| <i>A. racemosus</i>  | 0.004                   | 0.053         | 0.005             | 0.012             |
| <i>I. digitata</i>   | 0.007                   | 0.089         | 0.013             | 0.007             |
| <i>T. cordifolia</i> | 0.005                   | 0.076         | 0.002             | 0.015             |
| <i>W. somnifera</i>  | 0.008                   | 0.081         | 0.025             | 0.004             |

**3.6.2. Pesticide residual analysis**

The powdered mixture of polyherbal was subjected for the detection of pesticide residual analysis after purification by column chromatography and chromatogram is developed by GC-MS technique and detection is done based on the standard retention time of pesticides.

**Table: 7 Pesticide residual analysis**

| Parameters                 | Observation  |
|----------------------------|--------------|
| Organochlorine compound    | Not detected |
| Oragnophosphorous compound | Not detected |
| Carbamates                 | Not detected |

**3.6.3. Microbial load analysis**

Microbial screening for the raw materials was carried out and the results obtained were detailed in Table 8.

**Table 8: Microbial load analyses**

| Parameters                            | <i>A. paniculata</i> | <i>A. racemosus</i> | <i>I. digitata</i> | <i>T. cordifolia</i> | <i>W. somnifera</i> |
|---------------------------------------|----------------------|---------------------|--------------------|----------------------|---------------------|
| Total aerobic count (NMT 1000 cfu/g)  | NIL                  | 380cfu/g            | NIL                | 650cfu/g             | 650cfu/g20          |
| Yeast and mould count (NMT 100 cfu/g) | 53cfu/g              | NIL                 | 58cfu/g            | 28cfu/g              | NIL                 |
| <i>E.coli</i> (To be absent)          | absent               | absent              | absent             | absent               | absent              |
| <i>Salmonella</i> (To be absent)      | absent               | absent              | absent             | absent               | absent              |
| <i>Pseudomonas</i> (To be absent)     | absent               | absent              | absent             | absent               | absent              |
| <i>Staphylococcus</i> (To be absent)  | absent               | absent              | absent             | absent               | absent              |
| <i>Shigella</i> (to be absent)        | absent               | absent              | absent             | absent               | absent              |

### 3.7. Preformulation and Formulation Development Studies

Totally five trials of formulation were carried out using different choices of excipients considering different facets of

manufacturing problems as well as quality defects in mind, all the resultant formulations were evaluated for their flow property, uniformity of filling, uniformity of weight, moisture content and disintegration time.

**Table 9: Evaluation of trial batches**

| Parameters                          | Trial 1    | Trial 2   | Trial 3   | Trial 4   |
|-------------------------------------|------------|-----------|-----------|-----------|
| Bulk density (g/cm <sup>2</sup> )   | 0.42±0.01  | 0.38±0.05 | 0.35±0.04 | 0.33±0.06 |
| Tapped density (g/cm <sup>2</sup> ) | 0.54±0.03  | 0.61±0.02 | 0.53±0.03 | 0.57±0.02 |
| Compressibility index (%w/w/)       | 26.6±2.08  | 32±2.64   | 34.6±2.08 | 33.28±1.5 |
| Housner's ratio                     | 1.41±0.01  | 1.43±0.02 | 1.53±0.04 | 1.45±0.03 |
| Angle of repose (degrees)           | 43.03±3.78 | 37.04±1.0 | 35.05±1.0 | 27.6±0.57 |

Results are reported as Mean ± Standard deviation (n=3)

As per the standards, the flow property of the blend to be filled in the capsule should be in good range and was confirmed by the above parameters. Trial batch IV showed excellent flow characters and batch IV was taken for capsule filling.

The trial IV flow properties were Excellent and all parameters were within the Specified limits. So, fourth trial was chosen for further studies.

**Table 10: Evaluation of Inprocess Parameters**

| Parameter             | Trial I   | Trial II  | Trial III        | Trial IV         |
|-----------------------|-----------|-----------|------------------|------------------|
| Flow property         | Poor flow | Poor flow | Fair             | Good             |
| Uniformity of Filling | -         | -         | Uniform          | Uniform          |
| Uniformity of Weight  | -         | -         | Less weight      | Uniform          |
| Moisture content      | -         | -         | Within the limit | Within the limit |
| Disintegration time   | -         | -         | Within the limit | Within the limit |

Mean ± Standard Deviation (n=3)

### 3.8. Standardization of formulation

#### Capsule evaluation

##### Description

"light brown" coloured granules packed in "0" size blue capsules. The polyherbal capsules were evaluated for organoleptic characters which include colour, odour, taste and nature.

**Table 11: Organoleptic Characters of Capsules**

| Parameters  | Observation   |
|-------------|---|
| Description | Light brown granule in blue cap and body "0" size capsule |
| Colour      | Light brown granule                                       |
| Odour       | Characteristic odour                                      |
| Taste       | Bitter taste  |

**Table 12: Evaluation of capsules**

| Parameter               | Observation       |
|-------------------------|-------------------|
| Average weight          | 574.31 ± 4.5mg    |
| Weight variation        | Within I.P. Limit |
| Moisture content(LOD)   | 2.51±0.1 %w/w     |
| Disintegration time     | 10.9±0.5(min)     |
| pH(1% aqueous solution) | 5.52± 0.68        |

Result (n=3) are reported as Mean ± Standard deviation

### 3.9. Preliminary Phytochemical Screening of Capsules

**Table 13: Preliminary phytochemical screening**

| Phytoconstituents | Observation |
|-------------------|-------------|
| Alkaloid          | Present     |
| Flavanoid         | Present     |
| Tannin            | Present     |
| Saponin           | Present     |
| Glycoside         | Present     |
| Terpenoid         | Present     |
| Steroid           | Present     |
| Carbohydrate      | Present     |
| Phenolics         | Present     |

The results established a scientific data which can be used for the identification of the crude drugs.

### 3.10. Quantitative Estimation of Phytoconstituents

The Polyherbal formulation was found to contain various phytochemical constituents and hence it is desirable to quantify few of them in order to establish a standard to maintain its quality. Among them the estimation of total Alkaloids, phenolics, Flavanoids, and Tannin content in the aqueous extract were decided to be taken as parameters. Samples were

drawn from three random samples of polyherbal capsules and the total alkaloids, phenolics, Flavonoids, and Tannin content present in them were estimated.

The estimated amounts of Alkaloids, phenolics, Flavonoids, and Tannins were enumerated in the Table 1.

**Table 14: Quantitative estimation of phytoconstituents**

| Parameter               | Observation (% w/w) |
|-------------------------|---------------------|
| Total alkaloid content  | 1.57±0.58           |
| Total tannin content    | 0.54±0.15           |
| Total flavonoid content | 3.25±0.37           |
| Total phenolic content  | 1.75±0.21           |

Result (n=3) are reported as Mean ± Standard deviation

### 3.11. Fluorescence Analysis of Polyherbal Capsule

**Table 15: Fluorescence Analysis of Polyherbal Capsule**

| Sample                 | Before treatment |          |         | After treating with 50 % HCl |                |         | After treating with 50% NaOH |                 |            |
|------------------------|------------------|----------|---------|------------------------------|----------------|---------|------------------------------|-----------------|------------|
|                        | Ordinary light   | Short UV | Long UV | Ordinary light               | Short UV       | Long UV | Ordinary light               | Short UV        | Long UV    |
| Polyherbal formulation | Greenish brown   | Green    | Green   | Green                        | Greenish brown | Brown   | Greenish yellow              | Greenish yellow | Dark brown |

### 3.12. Microbial load analysis

Microbial screening is done for the Polyherbal formulation from the fresh packs of the Same Batch and the results obtained were detailed in Table 16.

**Table 16: Microbial load analyses**

| Parameter            | Result   | Limits as per WHO |
|----------------------|----------|-------------------|
| Total aerobic count  | 230cfu/g | NMT 1000 cfu/g    |
| Yeast and Mould      | 15cfu/g  | NMT 100 cfu/g     |
| <i>E.Coli</i>        | Absent   | Should be absent  |
| <i>Salmonella</i>    | Absent   | Should be absent  |
| <i>Pseudomonas</i>   | Absent   | Should be absent  |
| <i>Streptococcus</i> | Absent   | Should be absent  |
| <i>Shigella</i>      | Absent   | Should be absent  |

From the results, it is shown that the formulation complies with the WHO standards for Microbial load analysis and hence it is safer to be taken internally.

### 3.13. Chromatographic Finger Print Analysis



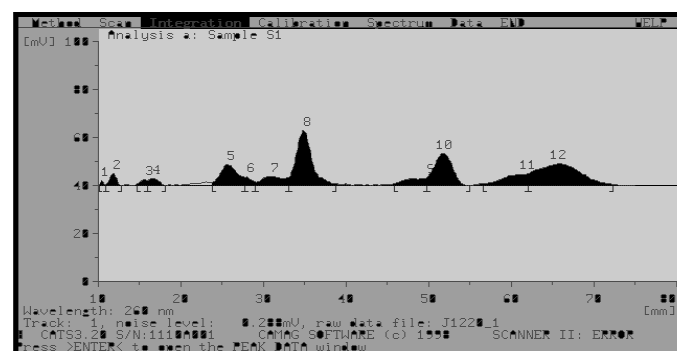
**Fig. 1: TLC photograph for Sample under 254nm**

### Mobile phase

Toluene:Ethylacetate:Methanol:Formicacid (6:3:1:0.2)

### Stationary phase

Aluminum coated Silica Gel – MerkF254



**Fig. 2: Chromatographic finger printing analysis of formulation**

**Table 17: Rf value of the capsules**

(Chromatographic finger printing)

| Rf   | Height | Area  | $\lambda$ max |
|------|--------|-------|---------------|
| 0.10 | 2.1    | 8.1   | 200           |
| 0.11 | 5.0    | 46.4  | 200           |
| 0.15 | 2.4    | 23.1  | 275           |
| 0.16 | 2.9    | 41.6  | 275           |
| 0.25 | 8.6    | 214.2 | 223           |
| 0.27 | 3.6    | 34.0  | 275           |
| 0.31 | 3.8    | 121.7 | 275           |
| 0.34 | 22.9   | 508.5 | 200           |
| 0.48 | 3.2    | 94.4  | 275           |
| 0.51 | 13.2   | 345.8 | 275           |
| 0.62 | 5.0    | 166.9 | 275           |
| 0.62 | 9.0    | 564.5 | 275           |

#### 4. CONCLUSION

In this study a Polyherbal capsule was formulated trail batch with the individually standardized raw materials of *Andrographis paniculata* (Stem and leaves), *Asparagus racemosus* (Root), *Ipomoea digitata* (Rhizome), *Tinospora cordifolia* (Stem) and *Withania somnifera* (Root) as per Ayurvedic Pharmacopoeia of India. The T4 was consider to a best batch as it complies with all the pharmacopoeial parameters and was selected for further study.

#### 5. REFERENCES

1. Huai H. *Ethnobot Res Appl.* 2010; **8**:169-79.
2. Husain SZ, Malik RN, Javaid M, Bibi S. *Pak J Bot.* 2008; **40**:1897-911.
3. Dewanjee S, Das AK, Sahu R, Gangopadhyay M. *Food Chem Toxicol* 2009, **47**:2679-2685.
4. Umar A, Ahmed QU, Muhammad BY, Dogarai BB, Soad SZ. *J Ethnopharmacol*, 2010, **1**:140-145.
5. Liu H, Liu X, Lee J, Liu Y, Yang H, Wang G. *Biochem Pharmacol*, **2008**(75):1649-1658.
6. Ministry of Health and Family Welfare. Ayurvedic Pharmacopoeia of India. 2011. Vol-VIII, P-193-195.
7. Harborne JB. *Phytochemical Methods: A Guide to modern techniques of plant analysis*. 2nd ed. London, Chapman and Hall 1973; p-434.
8. Kokate CK, Purohit AP, Gokahle SB. *Pharmacognosy*. 24th ed. Pune: Vallabh Prakashan; 2003; 108-9.
9. Lira, Sergio, Peter Brush, Laurence Senak, Chi San Wu, Edward Malawer. *Pharmacopoeial Forum*. 2008; **(3)**4:6-10.
10. Ministry of Health and Family Welfare. Ayurvedic Pharmacopoeia of India. 2008. Vol-IV, P-284.
11. Ministry of Health and Family Welfare. Ayurvedic Pharmacopoeia of India. 2008. Vol-IV, P-275-280.
12. The Theory and practice of industrial pharmacy by Leon Lachman Herbert A. Lieberman Joseph and Keing 3rd ed, published by Varghese publishing house, 2009, p-171-184
13. United States Pharmacopoeia. 30th ed. NF-25: The Official Standard of Compendia; 2007. Powder flow; p. 1174.
14. The Official Standard of Compendia; 2007. Bulk Density and Tapped Density; 30th ed. NF-25: p. 1186.
15. Ministry of health and family welfare. Indian pharmacopoeia. Ghaziabad: the Indian Pharmacopoeia Commission; 2007 vol2; p 76, 78, 134, 182, 191.
16. Fazel Shamsa, Hamiderza Monsef, Rouhollah Ghamooshi, Mohammadreza Verdian-riozi. *Thailand Journal of Pharmaceutical science*. 2008; **3**:17-20.
17. Edeiga HO, Okwu DE and Mbaebie BO. *African Journal of Biotechnology*. 2005; **4**(7):685-688.
18. Singleton VL, Orthofer R, Lamuela-Raventos RM. *Methods Enzymol*, 1999; **299**: 152-178.
19. Shivakumar BS. *Am. J. PharmTech Res.* 2012; **2**(5):417-422.
20. Gurdeep R, Chatwal, Sham K and Anand. *Instrumental methods of chemical analysis*. Mumbai: Himalaya publishing house; 2007. P.615.
21. Arun Paheed, Sravya reddy. B Roja.C. *International Journal of Phytotherapy*. 2012; **2**(2):74-88.